

100%; in the high-dose HNK treatment group, only 2 of 6 mice had lymphatic metastasis. The high-dose HNK also significantly decreased the weight of mediastinal lymph nodes over 80% compared with control group (Fig. 2C).

HNK inhibits metastasis of lung cancer cells to the brain *in vivo*

In this assay, we used an ultrasound-guided procedure to insure the injection of brain-seeking H2030-BrM3 lung cancer cells into the LV of NOD/SCID mice (Fig. 3A). One day after cell inoculation, the mice were randomly grouped to vehicle control and HNK low- (2 mg/kg b.w.) and high-dose (10 mg/kg b.w.) groups. High-dose HNK significantly decreased brain metastasis over 70% when compared with the vehicle control group (Fig. 3B). At necropsy (28 days post-LV injection), the extent of brain metastases was also quantified by *ex vivo* bioluminescence and GFP imaging as shown in Fig. 3C. HNK treatment decreased brain metastasis to approximately one third of that observed in control mice. Lung tumor cell migration to brain was confirmed by H&E

staining, as well as GFP staining (Fig. 3D and E). Collectively, our data suggest that HNK could be effective in preventing the metastasis of lung cancer cells to the brain.

STAT3 as a potential target of HNK in the inhibition of lung cancer brain metastasis

Potential mechanisms of action of HNK in the inhibition of lung cancer cell brain metastasis were examined via RTK assays (Fig. 4A). H2030-BrM3 cells were treated with 10 and 20 $\mu\text{mol/L}$ HNK for 6 hours. PathScan RTK signaling array revealed that HNK treatment dramatically decreased STAT3 phosphorylation (Fig. 4B and C), suggesting that STAT3 is at least one molecular target of HNK. The effects of HNK on STAT3 phosphorylation in PC9-BrM3 and H2030-BrM3 cells were confirmed by Western blot analysis (Fig. 4D). Previously, HNK was found to be effective in the treatment of head and neck squamous cell carcinoma (HNSCC) via targeting the EGFR signaling pathway (32). In the current study, we also observed that HNK targets the EGFR-AKT

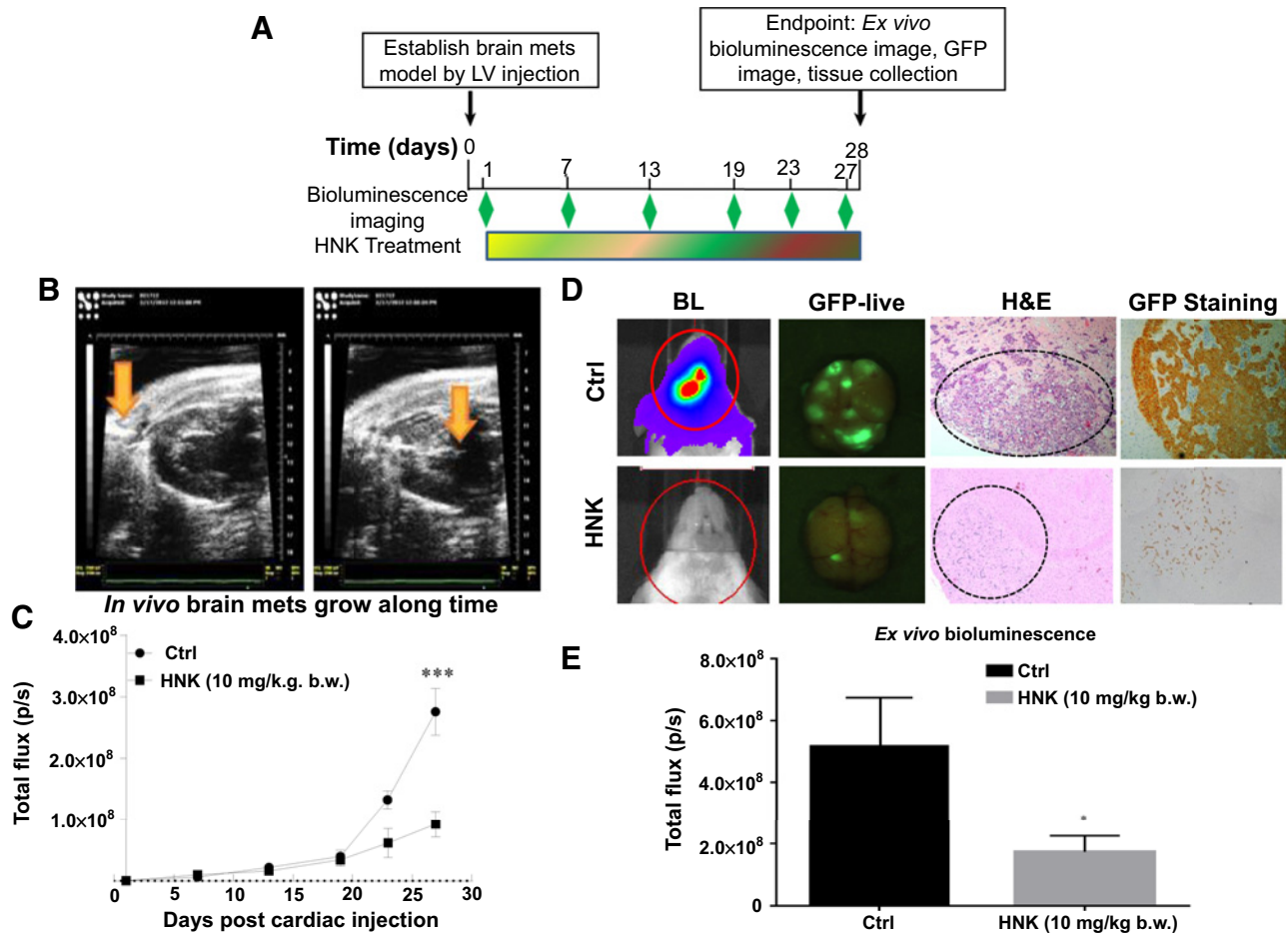
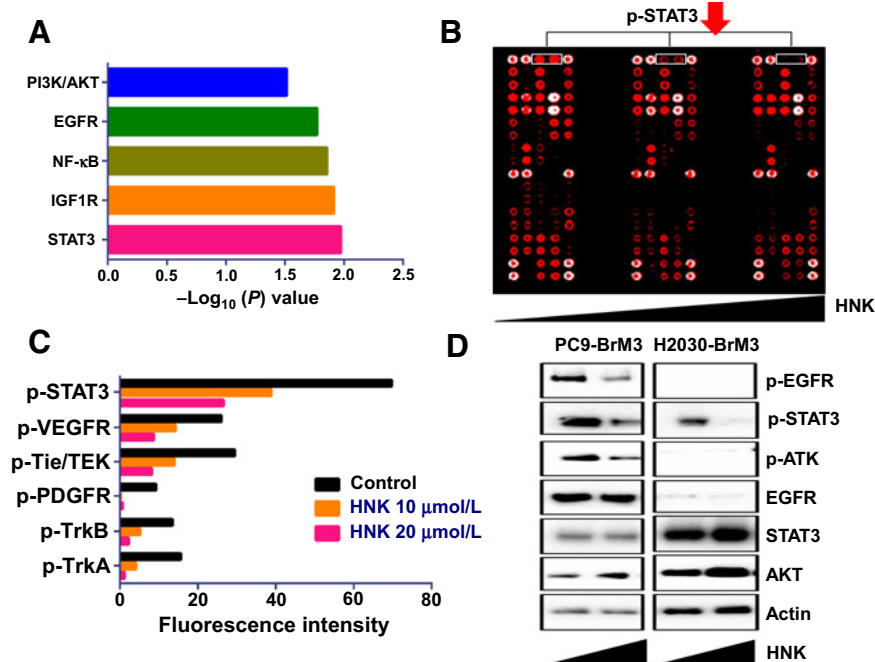


Figure 3.

HNK inhibits lung cancer brain metastasis. **A**, H2030-BrM3 cells expressing GFP and luciferase were engrafted in the arterial circulation by an ultrasound-guided LV injection. Brain metastasis was detected by bioluminescence at different time points with an IVIS 200 Xenogen monitor (Xenogen; exposure time, 1 minute; binning 8; no filter; f/stop16; field of view 12.5 cm). **B**, Corresponding grayscale photographs and color luciferase images are superimposed and analyzed with LivingImage (Xenogen). Mets, metastases. Data are expressed as normalized photon flux (photons/s/cm²). After final IVIS scan, mouse brain was dissected and imaged using Maestro Multi-Spectral Imaging System for GFP signal. **C**, Quantification of bioluminescence imaging signal intensity in the control (Ctrl) and HNK-treated group at different time points after the injection of H2030-BrM3 cells. Quantified values are shown in total flux. **D**, Representative luciferase, GFP, and H&E IHC images from mice treated with either gavage control (Ctrl) or HNK. **E**, Quantification of bioluminescence imaging signal intensity in the control (Ctrl) and HNK-treated group at the end of study via *ex vivo* Live Imaging. *, $P < 0.05$; ***, $P < 0.001$.

Pan et al.

**Figure 4.**

HNK targets STAT3 phosphorylation via inhibition of multiple RTKs. **A**, Pathway analysis based on RNA-seq revealed that HNK inhibits multiple RTK pathways, including EGFR, as well as the IGF1R and NF- κ B, PI3K/AKT, and STAT3 signaling pathway. Notably, STAT3 signaling pathway was the most significantly affected pathway by HNK. **B**, H2030-BrM3 cells were treated with HNK, and cell lysates were examined using RTK signaling arrays. STAT3 phosphorylation is downregulated by HNK in a dose-dependent manner. **C**, Effects of HNK on the phosphorylation of STAT3, as well as other signaling pathways, such as TrkA/B, PDGFR, Tie/TEK, and VEGFR signaling pathways. **D**, The phosphorylation of EGFR and AKT is downregulated by HNK only in the EGFR-mutant PC9-BrM3 cell line, but not in the Kras-mutant H2030-BrM3 cell line. STAT3 phosphorylation is downregulated by HNK in both cell lines.

signaling pathway in PC9-BrM3 cells, which harbor an EGFR mutation, but not in H2030-BrM3 cells, which harbor Kras mutations. In addition, we examined the interaction between HNK and multiple RTKs via the KINOMEScan binding assay. As shown in Supplementary Fig. S1, HNK did not bind directly to any of the RTKs tested.

STAT3 knockdown decreases the anticancer effects of HNK in lung cancer

shRNA knockdown was used to demonstrate the role of STAT3 in mediating the effects of HNK in lung cancer. Knockdown of STAT3 in PC9-BrM3 and H2030-BrM3 cells was validated by Western blot analysis (Fig. 5A). STAT3 knockdown decreases the antiproliferative (Fig. 5B) and antiinvasive (Fig. 5C and D) effects of HNK in both PC9-BrM3 and H2030-BrM3 cell lines. HNK treatment (20 μ mol/L) for 48 hours inhibited the proliferation of PC9-BrM3 vector control cells by 30% and H2030-BrM3 vector control cells by 20% but had no significant effect on the proliferation of STAT3 knockdown PC9-BrM3 or H2030-BrM3 cells (Fig. 5B). In addition, HNK treatment (10 μ mol/L) significantly inhibited the invasion of both PC9-BrM3 and H2030-BrM3 vector control cells but had no effect on the invasion of STAT3 knockdown PC9-BrM3 or H2030-BrM3 cells (Fig. 5C and D). Finally, we examined the effects of STAT3 on mitochondrial respiratory function in PC9-BrM3 and H2030-BrM3 cells. As shown in Fig. 5E, STAT3 knockdown significantly decreased mitochondrial respiratory function in both PC9-BrM3 and H2030-BrM3 cell lines. The anticancer effects of HNK, therefore, could be through inhibition of STAT3-mediated mitochondrial functions in lung cancer cells that have metastasized to the brain.

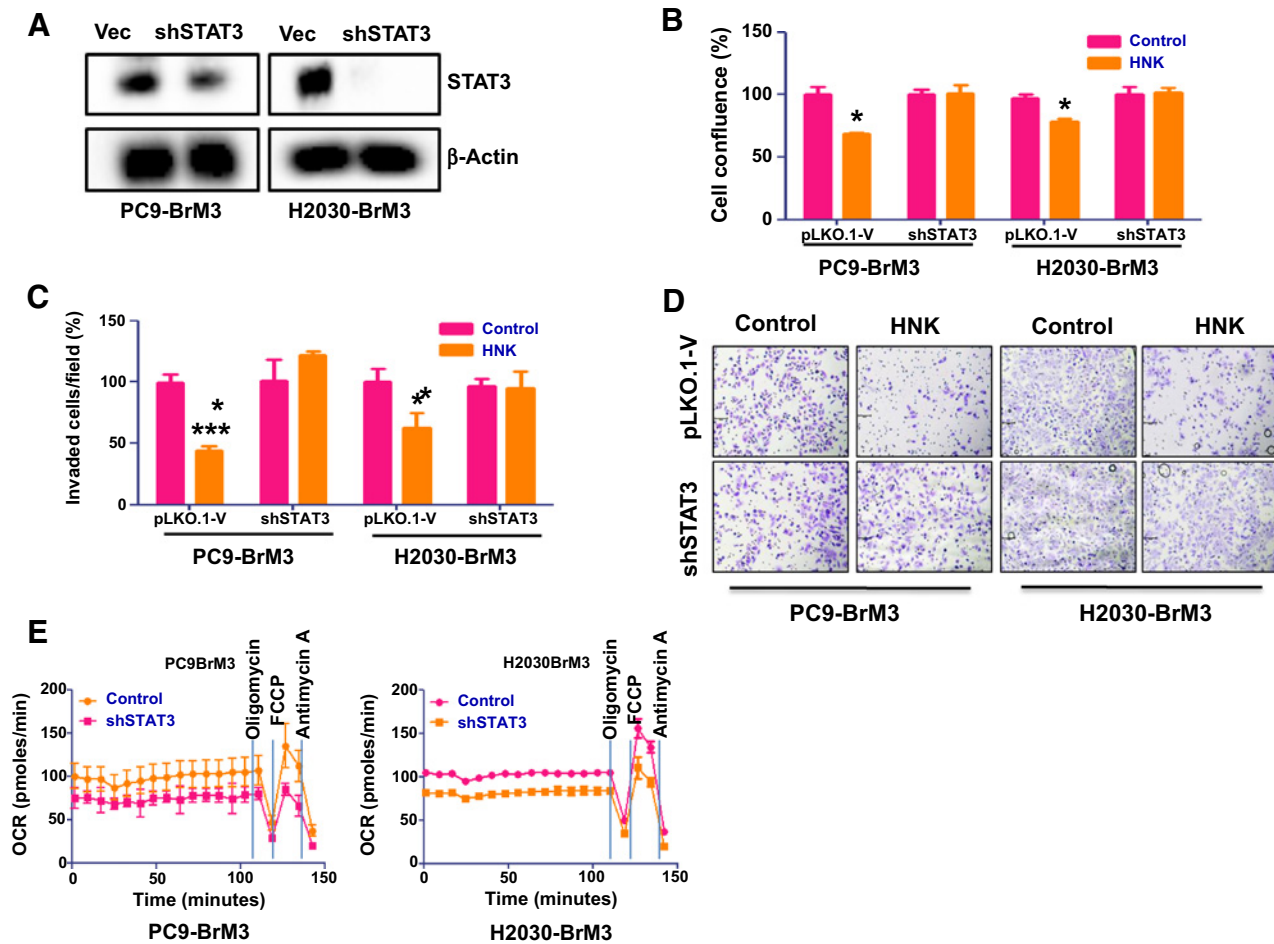
RNA-seq analysis showed that the expressions of key genes important to the activation of STAT3 pathway were downregulated in the metastatic lung tumors by HNK treatment

The differentially expressed genes identified by RNA-seq analysis software were subjected to IPA analysis (Ingenuity Pathway

Analysis; <http://www.ingenuity.com/products/ipa>) to identify the most significant oncogenic pathways in metastatic lung tumors changed by HNK treatment. Our genome-wide RNA-seq scan showed that STAT3 pathway is the top downregulated one among the oncogenic pathways that were significantly downregulated in the HNK-treated human lung tumor metastases in mouse brains (Fig. 4A). In addition, RNA-seq analyses identified that six key genes involved in the activation of STAT3 pathways were significantly downregulated in metastatic lung tumors *in vivo* upon HNK treatment (Table 1). They were FGFR4, IGF1R, IGF2R, MAP2K1, MAP3K11, and SRC. These matched the findings from our functional studies and supported that the anti-lung cancer role of HNK was mediated via the STAT3 signaling pathway.

Discussion

One of the common sites for metastases of lung cancers is the brain. Currently available therapies to address CNS metastases include whole brain/CNS irradiation or surgical resection in eligible patients, treatment with anti-EGFR agents in patients whose tumors contain EGFR mutations, as well as using next-generation ALK TKI that is brain-penetrable such as PF-06463922, to control CNS metastases in lung cancer patients (2, 33). However, these treatment options are available only after the diagnoses of brain metastases, and in many cases, metastatic lesions remain undiagnosed for long periods or they are not amenable to treatment with chemo/radiotherapy or surgery. Therefore, it is necessary to develop prevention strategies to inhibit metastases from primary tumors. Recently, we demonstrated the ability of HNK to potentially inhibit the development of lung tumors in mice (5). Analysis of HNK's mechanism of action suggests that its effect is primarily mediated by inducing apoptosis through a mitochondria-dependent mechanism (5, 7, 34). Here, through the use of the well-characterized brain metastases murine model, we report that HNK exerts inhibitory effects on the metastasis of lung cancer

**Figure 5.**

STAT3 knockdown abrogates the antiproliferative, antimigratory, and anti-invasive effects of HNK. **A**, Efficiency of STAT3 knockdown via shRNA approach in PC9-BrM3 and H2030-BrM3 was determined via Western blot analysis. The role of STAT3 in mediating the antiproliferative and anti-invasive effects was determined as indicated in Material and Methods. STAT3 knockdown abrogates the antiproliferative (**B**) and anti-invasive (**C**) effects of HNK in PC9-BrM3 and H2030-BrM3 cell lines. **D**, Effects of STAT3 in mitochondria respiratory function were examined via Seahorse experiment. STAT3 knockdown decreases the mitochondria respiratory function in both PC9-BrM3 and H2030-BrM3 cells. **E**, Representative images of PC9-BrM3 and H2030-BrM3 control vector-transfected and STAT3 knockdown cells with/without treatment of HNK from invasion assay.

cells to the brain, indicating that the compound has chemopreventive potential against both primary lung tumors and on metastasis of lung cancer to the brain.

Direct injection of tumor cells into the LV is the most widely used brain metastasis model in rodents because it bypasses the precolonization steps of dissemination of cancer cells through the bloodstream, homing, and extravasation, and recapitulates the process of cancer cells crossing the BBB and growing within the brain microenvironment. Metastatic brain lesions in mice vary from round, circumscribed lesions typical of that seen on human scans, to infiltrative tumor cells, which over time form typical round lesions that are ideal for evaluating the preventative effect of HNK on lung cancer metastasis. The brain homing H2030 and PC9 lung cancer cell lines were developed to have 100% brain metastatic potential (5–7, 29–31), H2030 cells with a KRAS^{G12C} mutation (35) and PC9 cell with an EGFR^{Δexon19} mutation (36). These cell lines were engineered to stably express GFP–luciferase fusion for real-time monitoring of metastatic tumor growth. In the

current study, we monitored metastatic tumor growth using both live animal imaging and endpoint *ex vivo* imaging. We also validated tumor growth by staining the brain tissues with H&E and GFP, and both stains consistently demonstrated about a 70% inhibition of brain metastases by HNK.

At least one of the mechanisms through which HNK inhibited lung cancer cell metastasis to the brain was through the inhibition of STAT3 phosphorylation. HNK is known to target multiple signaling pathways, including EGFR, MAPK, and PI3K/AKT (10–13). Recently, Sirt3 and GRP78 were also suggested as potential binding targets of HNK in different tissue types (34, 37). Interestingly, STAT3 is a major downstream mediator of multiple RTK pathways (14–17). Our data suggest that STAT3 could be a universal downstream target of HNK treatment. As indicated before, HNK was effective in the treatment of HNSCC via targeting the EGFR signaling pathway (38). The brain homing H2030 and PC9 lung cancer cell lines carry different driver mutations, H2030 with a KRAS^{G12C} mutation (35) and PC9 with

Table 1. Characteristics of the six key genes in the STAT3 pathway that was downregulated in the HNK-treated human lung tumor metastases in mouse brains

Symbol	Entrez gene name	Fold change (HNK vs. non-HNK Mets)	FDR	Location	Type(s)
<i>FGFR4</i>	Fibroblast growth factor receptor 4	-18.5	0.040	Plasma membrane	Kinase
<i>IGF1R</i>	Insulin-like growth factor 1 receptor	-2.3	0.022	Plasma membrane	Transmembrane receptor
<i>IGF2R</i>	Insulin-like growth factor 2 receptor	-1.6	0.012	Plasma membrane	Transmembrane receptor
<i>MAP2K1</i>	Mitogen-activated protein kinase kinase 1	-1.7	0.011	Cytoplasm	Kinase
<i>MAP3K1</i>	Mitogen-activated protein kinase kinase kinase 11	-1.8	0.011	Cytoplasm	Kinase
<i>SRC</i>	SRC proto-oncogene, non-receptor tyrosine kinase	-2.0	0.003	Cytoplasm	Kinase

Abbreviation: Mets, metastases.

an EGFR^{Δexon19} mutation (36). In PC9-BrM3 cells, we observed downregulation of phosphorylated EGFR by HNK, but not in H2030-BrM3 cells, which do not carry an EGFR mutation. Therefore, the effects of HNK on the EGFR-AKT signaling pathway could be cell type- or tissue-specific. RNA-seq data suggest that *FGFR4* is the most significant gene that was affected by HNK treatment, and *FGFR4* is known to mediate STAT3 signaling pathway (39). Therefore, it will be interesting to investigate the role of *FGFR4*-STAT3 signaling pathway in mediating the anti-cancer effects of HNK. STAT3 phosphorylation was reduced in both PC9-BrM3 and H2030BrM lung cancer cell lines by HNK, and knockdown of endogenous STAT3 in these cell lines abrogated the antiproliferative, antimigratory, and anti-invasive effects of HNK, further supporting the concept that STAT3 could be a universal downstream target of HNK regardless of EGFR mutation status of lung cancer cells. Although, with prolonged treatment (over 48 h), HNK will eventually inhibit proliferation of STAT3 knockdown cells (data not shown), which most likely would be due to the off-target effects of HNK, considering it is a polyphenol compound, once its main target has been blocked, it may target other pathways to inhibit tumor growth. STAT3 knocked down PC9-BrM3 and H2030-BrM3 cells exhibit significantly less mitochondrial respiratory function than normal lung cells. HNK inhibition of lung cancer progression via inhibition of mitochondrial respiratory function could, therefore, be due to inhibition of STAT3 phosphorylation which, in turn, leads to inhibition of the metastases of lung cancer cells to the brain.

References

- Jemal A, Siegel R, Xu J, Ward E. Cancer statistics, 2010. *CA Cancer J Clin* 2010;60:277-300.
- Goldberg SB, Contessa JN, Omay SB, Chiang V. Lung cancer brain metastases. *Cancer J* 2015;21:398-403.
- Tsai TH, Chou CJ, Cheng FC, Chen CF. Pharmacokinetics of honokiol after intravenous administration in rats assessed using high-performance liquid chromatography. *J Chromatogr B Biomed Appl* 1994; 655:41-5.
- Chen F, Wang T, Wu YF, Gu Y, Xu XL, Zheng S, et al. Honokiol: a potent chemotherapy candidate for human colorectal carcinoma. *World J Gastroenterol* 2004;10:3459-63.
- Pan J, Zhang Q, Liu Q, Komasa SM, Kalyanaraman B, Lubet RA, et al. Honokiol inhibits lung tumorigenesis through inhibition of mitochondrial function. *Cancer Prev Res* 2014;7:1149-59.
- Wang X, Duan X, Yang C, Zhang X, Deng L, Zheng H, et al. Honokiol crosses BBB and BCSFB, and inhibits brain tumor growth in rat 9L intracerebral gliosarcoma model and human U251 xenograft glioma model. *PLoS One* 2011;6:e18490.
- Lin JW, Chen JT, Hong CY, Lin YL, Wang KT, Yao CJ, et al. Honokiol traverses the blood-brain barrier and induces apoptosis of neuroblastoma cells via an intrinsic bax-mitochondrion-cytochrome c-caspase pathway. *Neuro Oncol* 2012;14:302-14.
- Chen YJ, Wu CL, Liu JF, Fong YC, Hsu SF, Li TM, et al. Honokiol induces cell apoptosis in human chondrosarcoma cells through mitochondrial dysfunction and endoplasmic reticulum stress. *Cancer Lett* 2010;291: 20-30.
- Hahm ER, Singh SV. Honokiol causes G0-G1 phase cell cycle arrest in human prostate cancer cells in association with suppression of retinoblastoma protein level/phosphorylation and inhibition of E2F1 transcriptional activity. *Mol Cancer Ther* 2007;6:2686-95.
- Garcia A, Zheng Y, Zhao C, Toschi A, Fan J, Shraibman N, et al. Honokiol suppresses survival signals mediated by Ras-dependent phospholipase D activity in human cancer cells. *Clin Cancer Res* 2008; 14:4267-74.
- Crane C, Panner A, Pieper RO, Arbiser J, Parsa AT. Honokiol-mediated inhibition of PI3K/mTOR pathway: a potential strategy to overcome immunoresistance in glioma, breast, and prostate carcinoma without impacting T cell function. *J Immunother* 2009;32:585-92.
- Tse AK, Wan CK, Shen XL, Yang M, Fong WF. Honokiol inhibits TNF-α-stimulated NF-κB activation and NF-κB-regulated gene expression through suppression of IKK activation. *Biochem Pharmacol* 2005;70: 1443-57.
- Deng J, Qian Y, Geng L, Chen J, Wang X, Xie H, et al. Involvement of p38 mitogen-activated protein kinase pathway in honokiol-induced apoptosis in a human hepatoma cell line (hepG2). *Liver Int* 2008;28:1458-64.
- Wu J, Patmore DM, Jousma E, Eaves DW, Breving K, Patel AV, et al. EGFR-STAT3 signaling promotes formation of malignant peripheral nerve sheath tumors. *Oncogene* 2014;33:173-80.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Authors' Contributions

Conception and design: Y. Lee, M. You

Development of methodology: J. Pan, Q. Zhang, T.C. Wan

Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): J. Pan, Y. Lee, Q. Zhang

Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): J. Pan, Y. Lee, Q. Zhang, D. Xiong

Writing, review, and/or revision of the manuscript: J. Pan, Y. Lee, Q. Zhang, D. Xiong, Y. Wang, M. You

Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): Y. Lee, Y. Wang

Study supervision: M. You

Acknowledgments

We thank Dr. John Auchampach for his contribution, especially for the guidance of the left ventricle under ECHO 707.

Grant Support

This work was supported by R01CA208648.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received May 13, 2016; revised September 24, 2016; accepted November 1, 2016; published OnlineFirst November 14, 2016.

15. De Simone V, Franze E, Ronchetti G, Colantoni A, Fantini MC, Di Fusco D, et al. Th17-type cytokines, IL-6 and TNF-alpha synergistically activate STAT3 and NF-kB to promote colorectal cancer cell growth. *Oncogene* 2015;34:3493-503.
16. Zhou J, Wulfkuhle J, Zhang H, Gu P, Yang Y, Deng J, et al. Activation of the PTEN/mTOR/STAT3 pathway in breast cancer stem-like cells is required for viability and maintenance. *Proc Natl Acad Sci U S A* 2007;104:16158-63.
17. Yau CY, Wheeler JJ, Sutton KL, Hedley DW. Inhibition of integrin-linked kinase by a selective small molecule inhibitor, QLT0254, inhibits the PI3K/PKB/mTOR, Stat3, and FKHR pathways and tumor growth, and enhances gemcitabine-induced apoptosis in human orthotopic primary pancreatic cancer xenografts. *Cancer Res* 2005;65:1497-504.
18. Zhang Q, Raje V, Yakovlev VA, Yacoub A, Szczepanek K, Meier J, et al. Mitochondrial localized Stat3 promotes breast cancer growth via phosphorylation of serine 727. *J Biol Chem* 2013;288:31280-8.
19. Lin L, Liu A, Peng Z, Lin HJ, Li PK, Li C, et al. STAT3 is necessary for proliferation and survival in colon cancer-initiating cells. *Cancer Res* 2011;71:7226-37.
20. Yang H, Yamazaki T, Pietrocola F, Zhou H, Zitvogel L, Ma Y, et al. STAT3 inhibition enhances the therapeutic efficacy of immunogenic chemotherapy by stimulating type 1 interferon production by cancer cells. *Cancer Res* 2015;75:3812-22.
21. Langmead B, Trapnell C, Pop M, Salzberg SL. Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. *Genome Biol* 2009;10:R25.
22. Trapnell C, Pachter L, Salzberg SL. TopHat: discovering splice junctions with RNA-Seq. *Bioinformatics* 2009;25:1105-11.
23. Anders S, Pyl PT, Huber W. HTSeq—a Python framework to work with high-throughput sequencing data. *Bioinformatics* 2015;31:166-9.
24. Robinson MD, McCarthy DJ, Smyth GK. edgeR: a Bioconductor package for differential expression analysis of digital gene expression data. *Bioinformatics* 2010;26:139-40.
25. Bradford JR, Farren M, Powell SJ, Runswick S, Weston SL, Brown H, et al. RNA-seq differentiates tumour and host mRNA expression changes induced by treatment of human tumour xenografts with the VEGFR tyrosine kinase inhibitor cediranib. *PLoS One* 2013;8:e66003.
26. Rossello FJ, Tothill RW, Britt K, Marini KD, Falzon J, Thomas DM, et al. Next-generation sequence analysis of cancer xenograft models. *PLoS One* 2013;8:e74432.
27. Nguyen DX, Chiang AC, Zhang XH, Kim JY, Kris MG, Ladanyi M, et al. WNT/TCF signaling through LEF1 and HOXB9 mediates lung adenocarcinoma metastasis. *Cell* 2009;138:51-62.
28. Arora S, Singh S, Piazza GA, Contreras CM, Panyam J, Singh AP. Honokiol: a novel natural agent for cancer prevention and therapy. *Curr Mol Med* 2012;12:1244-52.
29. Arora S, Bhardwaj A, Srivastava SK, Singh S, McClellan S, Wang B, et al. Honokiol arrests cell cycle, induces apoptosis, and potentiates the cytotoxic effect of gemcitabine in human pancreatic cancer cells. *PLoS One* 2011;6:e21573.
30. Nagalingam A, Arbiser JL, Bonner MY, Saxena NK, Sharma D. Honokiol activates AMP-activated protein kinase in breast cancer cells via an LKB1-dependent pathway and inhibits breast carcinogenesis. *Breast Cancer Res* 2012;14:R35.
31. Singh T, Katiyar SK. Honokiol inhibits non-small cell lung cancer cell migration by targeting PGE(2)-mediated activation of beta-catenin signaling. *PLoS One* 2013;8:e60749.
32. Park EJ, Min HY, Chung HJ, Hong JY, Kang YJ, Hung TM, et al. Down-regulation of c-Src/EGFR-mediated signaling activation is involved in the honokiol-induced cell cycle arrest and apoptosis in MDA-MB-231 human breast cancer cells. *Cancer Lett* 2009;277:133-40.
33. Awad MM, Shaw AT. ALK inhibitors in non-small cell lung cancer: crizotinib and beyond. *Clin Adv Hematol Oncol* 2014;12:429-39.
34. Martin S, Lamb HK, Brady C, Lefkove B, Bonner MY, Thompson P, et al. Inducing apoptosis of cancer cells using small-molecule plant compounds that bind to GRP78. *Br J Cancer* 2013;109:433-43.
35. Phelps RM, Johnson BE, Ihde DC, Gazdar AF, Carbone DP, McClintock PR, et al. NCI-navy medical oncology branch cell line data base. *J Cell Biochem Suppl* 1996;24:32-91.
36. Koizumi F, Shimoyama T, Taguchi F, Saijo N, Nishio K. Establishment of a human non-small cell lung cancer cell line resistant to gefitinib. *Int J Cancer* 2005;116:36-44.
37. Pillai VB, Samant S, Sundaresan NR, Raghuraman H, Kim G, Bonner MY, et al. Honokiol blocks and reverses cardiac hypertrophy in mice by activating mitochondrial Sirt3. *Nat Commun* 2015;6:6656.
38. Singh T, Gupta NA, Xu S, Prasad R, Velu SE, Katiyar SK. Honokiol inhibits the growth of head and neck squamous cell carcinoma by targeting epidermal growth factor receptor. *Oncotarget* 2015;6:21268-82.
39. Tateno T, Asa SL, Zheng L, Mayr T, Ullrich A, Ezzat S. The FGFR4-G388R polymorphism promotes mitochondrial STAT3 serine phosphorylation to facilitate pituitary growth hormone cell tumorigenesis. *PLoS Genet* 2011;7:e1002400.

Cancer Prevention Research

Honokiol Decreases Lung Cancer Metastasis through Inhibition of the STAT3 Signaling Pathway

Jing Pan, Yongik Lee, Qi Zhang, et al.

Cancer Prev Res 2017;10:133-141. Published OnlineFirst November 14, 2016.

Updated version	Access the most recent version of this article at: doi:10.1158/1940-6207.CAPR-16-0129
Supplementary Material	Access the most recent supplemental material at: http://cancerpreventionresearch.aacrjournals.org/content/suppl/2016/11/12/1940-6207.CAPR-16-0129.DC1

Cited articles	This article cites 39 articles, 8 of which you can access for free at: http://cancerpreventionresearch.aacrjournals.org/content/10/2/133.full#ref-list-1
-----------------------	---

E-mail alerts	Sign up to receive free email-alerts related to this article or journal.
Reprints and Subscriptions	To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org .
Permissions	To request permission to re-use all or part of this article, use this link http://cancerpreventionresearch.aacrjournals.org/content/10/2/133 . Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.